

Original Articles**HEMOGLOBIN E-BETA THALASSEMIA IN UTTAR PRADESH****Sarita Agarwal, Reena Gulati and Kuldeep Singh**

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Objective: To evaluate the molecular make up of hemoglobin E-Beta thalassemia to facilitate diagnosis, genetic counseling and prenatal diagnosis in Uttar Pradesh. Design: DNA analysis. Setting: Referred hemolytic anemia cases to Genetics OPD of a tertiary care center. Subjects: 21 families of HbE-thalassemia of which 19 were of UP origin. Methods: The patient and obligate carriers in their families were evaluated at hematological, biochemical and molecular level. A total of 62 cases were evaluated which included the index cases and their family members. Red blood cell indices, osmotic fragility, hemoglobin electrophoresis, quantitation of fetal hemoglobin, HbA2/E, serum iron and total iron binding capacity estimation were carried out in all the blood samples. DNA analysis was done for HbE and beta thalassemia mutations. Results: The commonest, IVSI-5 (G→C) mutation (57%) was found along with HbE mutation. Only 23/26 cases belonged to the group of common P-thal mutations as described in literature. Conclusion: Establishment of antenatal diagnostic services is necessary in those parts of India where both these mutations are commonly seen.

Key words: Amplification refractory mutation system, Antenatal diagnosis, beta thalassemia, Hemoglobin E, Mutations.

HEMOGLOBIN E-Beta thalassemia is a disorder with a high frequency in South-East Asia and has been reported from different parts of India(1-14). Various factors decide the outcome of the interplay between this combination of a qualitative and a quantitative defect of hemoglobin. The clinical picture produced by the interaction is heterogeneous. We report here 26 patients with HbE thalassemia disease from 21 families where parents are obligate heterozygotes for HbE and **p-thal** mutations. An effort has been made to characterize the molecular make up of these patients

with a view to develop antenatal services for such families.

Subjects and Methods

Twenty six individuals from 21 families were diagnosed to be cases of [3-thalassemia HbE interaction. These patients had been referred to the Genetics OPD from 1987-1995 for investigation of refractory anemia. The index cases and available family members were clinically evaluated and investigated for the cause of anemia.

Patients Groups

Red blood cell indices were measured on automated Sysmax 800. Osmotic fragility was calculated according to the test described by Dacie and Lewis(15). Fetal hemoglobin (HbF) was estimated by alkali denaturation technique(16) and also by the methodology of Singers *et al.*(17). Quantitative estimation of hemoglobin A₂ (HbA₂) was carried out by column chromatography(18). Prepared hemolysates were subjected to Hb electrophoresis on cellogel at pH 8.5 in tris-glycine buffer for 90 minutes. DNA was extracted from peripheral blood leucocytes(19). Amplification refractory mutation system (ARMS) technique(20) was applied to confirm the presence of HbE mutation and to characterize the (3-thalassemia mutations.

Following primers were used to detect the presence of HbE:

5' ACC TCA CCC TGT GGA GCC AC - *

5' AA CC TG CCC AGG GGC TT <-

Primers used for detection of the 10 commonest (3-thalassemia mutations found in Indians were the same as described earlier(21,22).

ARMS

The basis of the system is the observation that oligonucleotides that are complementary to a given DNA sequence, except for a mismatch at their 3'-OH residue, will not function as primers in the PCR under appropriate conditions. A typical ARMS test consists of two complementary reactions, one primer for specific normal sequence and another for specific mutant sequence. The genotype of an individual can be determined by analysis of the amplification products. A normal individual generates PCR products only in the normal reaction, a heterozygote gives products in both reactions, and a homozygote mutant individuals does so only in the mutant reaction.

Results

Of the 21 families with 26 affected individuals, 19 belonged to Uttar Pradesh and one each to Bihar and Haryana. These included one Christian, 2 Muslim and 18 Hindu families. The patients were placed in 2 groups on the basis of age at onset and requirement for regular blood transfusions. Patients with milder phenotype, later age of onset, without regular need for blood transfusions were placed in Group A. Group B consisted of patients with severe anemia and early age of onset who required regular blood transfusions to maintain Hb levels beyond 6g/dl, except for one 18 year old girl who became symptomatic at 8-9 year of age but now requires regular blood support. The age range in this group was 2-18 years with 7 out of 11 patients aged below 5 years. In contrast, the age range in Group A was 4-38 years with 12 of the 15 patients being above 5 years and 9 out of 15 subjects older than 10 years of age.

The hematological parameters of the two groups are summarized in *Table I*. The mean Hb levels in Groups A and B were 7.9 and 5.0 g/dl, respectively. The mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) were similar in the two groups. Osmotic fragility (OF) was reduced in all patients to < 80% at 0.36% saline, except for two patients, one in each group.

HbE percentage was significantly higher in Group A with a mean of 53.9% and range of 24.6-81.0%. Group B showed a mean value of 41.2% with a range of 17.8-57.9%. However, the Group B patients with a severe phenotype had slightly higher mean HbF level of 25.1% as compared to 22.0% in Group A. The implication is difficult to interpret due to the small sample size.

TABLE Y-Hemalogical, Biochemical and Molecular Comparison of HbE-Beta Thalassemia Families.

	S. No.	Age/ Sex	Hb (g/dl)	MCV (fe)	MCH (Pg)	MCHC (g/dl)	OF (%)	HbE/A2 (%)	HbF (%)	E/p-thal
<i>Group A:</i>	1.	29/M	7.6	65.1	17.3	26.5	45.7	81.0	10.6	E/?
<i>Mild</i>	2.	6/M	7.3	55.0	-	-	39.0	55.7	26.6	E/VIS1-5 (G->C)
	3.	4/F	10.0	76.9	20.8	27.0	33.0	24.6	3.3	E/Co30 (G-^C)
	4.	12/M	10.8	63.9	24.3	38.2	-	41.8	30.1	E/IVS 1-5 (G->C)
	5.	34/F	7.9	61.0	16.2	26.0	44.0	59.0	18.4	E/IVS1-5 (G->C)
	6.	22/F	8.1	54.0	20.2	36.8	61.0	47.3	31.4	E/IVS 1-5 (G-^C)
	7.	34/M	7.2	63.0	19.5	31.0	42.4	69.9	19.0	E/CO 15 (G->A)
	8.	30/F	6.9	73.0	20.2	29.2	83.0	54.2	20.3	E/IVSI-5 (G->C)
	9.	7/M	7.1	70.6	-	-	15.4	52.0	23.7	E/Co30 (G->C)
	10.	5/M	7.3	57.9	-	-	24.8	60.0	12.5	E/Co30 (G->C)
	11.	19/M	9.2	78.0	22.0	30.0	48.0	51.8	22.6	E/Co 15 (G->A)
	12.	11/M	7.3	71.0	19.4	27.3	59.1	60.9	19.4	E/FSH 41-42
	13.	9/M	7.9	74.9	21.9	29.2	72.2	35.6	45.6	E/FSH 41-42
	14.	20/M	6.9	60.0	17.1	28.9	38.7	52.9	23.6	E/?
	15.	38/M	7.9	62.0	16.2	25.6	41.0	63.8	23.1	E/?
Mean		--	7.9	65.7	19.6	29.6	46.2	53.9	22.0	
<i>Group B:</i>	1.	3/M	5.4	68.0	21.3	31.4	37.4	39.9	29.9	E/Co 16 (-C)
<i>Severe</i>	2.	9/M	5.2	61.5	-	-	55.5	52.0	23.8	E/IVSI-5 (G->C)
	3.	7/F	5.6	69.5	21.6	35.0	55.6	46.8	22.0	E/IVSI-5 (G->C)
	4.	2.5/F	5.6	62.3	14.2	23.0	62.0	17.8	24.0	E/IVSI-5 (G->C)
	5.	9/M	5.6	60.0	14.0	23.3	65.6	34.2	31.3	E/IVSI-5 (G->C)
	6.	3/F	3.0	77.5	-	-	51.0	57.9	15.6	E/Co 15 (G->A)
	7.	2/M	5.6	77.0	24.0	31.1	-	19.2	7.0	E/Co 15 (G->A)
	8.	4/M	4.8	63.0	18.3	30.0	73.6	48.3	37.4	E/IVS I-5(G->C)
	9.	3/M	5.0	63.0	20.8	33.3	74.2	42.6	43.3	E/IVS 1-5 (G->C)
	10.	5/M	4.6	44.5	20.7	32.1	58.0	46.1	5.1	E/IVS 1-5 (G->C)
	11.	18/F	4.6	57.0	20.8	26.7	86.7	48.2	36.8	E/IVSI-5 (G->C)
Mean		-	5.0	63.9	19.5	29.5	62.0	41.2	25.1	

Presence of HbE mutation was confirmed in all patients, on analysis by ARMS. The (i-thalassemia mutation could only be characterized in 12 of the 15 patients in Group A and all in Group B with a panel of probes for the 10 most commonly found mutations in India (Fig. 1).

All the 8/11 (73%) characterized muta-

tions in Group B were IVS-I, 5 (G->C), whereas this was present in only 5/12 (42%) characterized mutations in Group A. Two siblings from Group A revealed a frameshift 41-42 (-CTTT) mutation. One was with Col6 in Group B and two each with Col5 from both groups were identified. However, three cases were found to have an uncommon mutation Co30 (G->C)

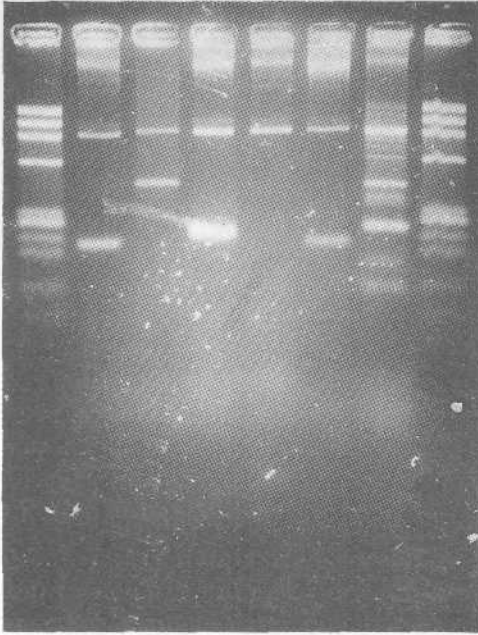


Fig. 1 *Ethidium bromide stained PCR products on 2% agarose gel. Lane 1 - Hae. III digested $\times 174$ DNA Molecular weight marker. It yields fragments of 1353, 1078, 872, 603, 310, 281, 234, 194 and 118 bp. Lane 2 & 6-861 and 239 bp fragments corresponding to Codon 16 (-c) mutation. Lane 3-861 and 443 bp fragments corresponding to Frame shift codon 41-42 (-CTTT) mutation. Lane 4-861 and 285 bp fragments corresponding to IVS 1-5 (G-C) mutation. Lane 5-861 bp fragment in normal sample used as positive internal control for PCR amplification.*

which was detected by DNA sequencing in Dr. Hattori's Laboratory, (α -thalassemia mutations in other three patients of Group A could not be characterized by the panel of probes for these 10 mutations.

Discussion

The heterogeneity in clinical and hematological picture of HbE/ α -thalassemia interaction is apparent from the results. Those with severe anemia and early

age of onset have a disease course similar to homozygous α -thalassemia while those with milder anemia and later age of onset get away with occasional need for blood transfusion or completely asymptomatic course of disease, hemolytic facies being the only indicator to presence of disease.

High HbF values are known to reduce the severity of symptoms. In our patients, however the HbF levels were not very different in the two groups. Hence the factor responsible for milder phenotype may be different from the protective values of HbF.

The percentage and range of HbE is significantly higher in Group A and is probably one of the factors responsible for the mild symptoms of these patients. HbE is a mild structural variant of β (3) globin chain, being asymptomatic in homozygous state. The severity of phenotype is, therefore, dependent on the type of β (3) thai mutation, HbE and HbF levels and number of α -globin genes which tend to reduce the severity of disease by altering the ratio of imbalance of α and β (3)-globin chains. The two α -gene deletions in association of β (3) type could be of less severe type as compared to either none or one α -gene deletion in association with β (12,23,24).

The interaction of genes for α and (β -thal, HbE, Hb-constant spring and other mutations results in a complex variety of phenotypes(25-27). A remarkable high incidence of deletion and non-deletion type of α -thalassemia mutants and its interaction with β -thal have been found in Indians(27,28). However, in our patients interplay of α -thalassemia genes possibly affecting the EB^T phenotype, has not been ruled out.

The commonest thalassemia mutation in our patients was IVS-1,5 (G->C). Surprisingly, only 23/26 mutations belonged to the group of 10 most commonly found

(3-thal mutations responsible for 97-98% of all cases of (J- thai in India. The 8 mutations characterized in Group B and 5 in Group A were IVS-1,5 (G->C). This is a B⁺ mutation and is probably contributory to the severity of the phenotype.

One family with two affected children (Christian) showed a frameshift mutation [FSC 41-42-(CTTT)]. Three patients were identified to be carrying an uncommon mutation, Co 30 G->C. However, 4 cases were carrying mutation at Col5 and only in one case Col6 mutation was identified. Thus only 4 of the 10 commonest mutations contributed to our 26 cases of E/p thai interaction.

Our plan is to try and further characterize the remaining 3 undiagnosed (J thai mutations in these patients. Molecular diagnosis of HbE is required for first trimester antenatal diagnosis besides characterization of P-thal mutation. An establishment of this kind of antenatal diagnostic services is the ultimate aim of the study.

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REFERENCES

- Pande SR, Mehrotra VG, Mehrotra TN. Study of abnormal hemoglobins in professional blood donors. *Indian J Med Res* 1972; 58: 283-284.
- Gupta SC, Mehrotra TN, Sharma NP, *et al.* Abnormal hemoglobins in Nepali Gorkhas. *Indian J Med Res* 1977; 66: 809-814.
- Gupta SC, Mehrotra TN, Mehrotra VG. HbE thalassemia in Uttar Pradesh. *Indian J Med Res* 1970; 58: 857-862.
- Chatterjee JB. Hemoglobinopathy in India. *In: Abnormal Hemoglobin*. Eds. Jonxis JHP, Delafresnaye JF. Oxford, Blackwell Scientific Publications, 1959; p 322.
- Chatterjee JB. Some aspects of HbE and its genetic interaction with thalassemia. *Indian J Med Res* 1965; 53: 377-383.
- Chatterjee JB, Swarup S, Ghosh SK, *et al.* Incidence of HbE and of thai trait in Bengalis. *Bull Cal Sch Trop Med* 1957; 5: 159-160.
- Desai AJ, Bichile SK, Mehta NB, *et al.* Clinical profile of thai syndromes in India. *Birth Defects* 1988; 23 (5A): 275-280.
- Sukumaran PK, Landelia HP, Sanghvi LD, *et al.* Thalassemia syndrome in Bombay. *J Assoc Phys India* 1961; 9: 477-481.
- Mehrotra VG, Gupta SC, Pande SR, *et al.* Abnormal hemoglobins in Uttar Pradesh. *Indian J Med Res* 1968; 56:1365-1370.
- Sharma NP, Gupta SC, Atal PR, *et al.* Abnormal hemoglobins in Pakistani Armed Forces Personnel. *Indian J Med Res* 1976; 64: 883-890.
- Sharma RS, Parekh JG. Hemoglobinopathies in Western India. *Annual Report Indian Society of Hematology* 1963; 4: 36-42.
- Weatherall DJ, Clegg JB. *The Thalassemia Syndrome*, 3rd edn. Oxford, Blackwell Scientific Publication, 1981; p 303.
- Pati AR, Bhargava M, Rath PK, *et al.* Unusual features of HbE-thalassemia. *Indian J Med Res* 1985; 81: 409-412.
- Mathur KS, Mehrotra TN, Dayal RS, *et al.* Incidence of HbE and thalassemia in UP. *J Indian Med Assoc* 1962; 39:172-177.

15. Dacie JV, Lewis SM. Investigations of abnormal hemoglobins and thalassemias. *In: Practical Hematology*, 7th edn Edinburgh, Churchill Livingstone, 1991; p 227.
16. Betke K, Marti HR, Schlicht I. Estimation of small percentage of fetal hemoglobin. *Nature* 1959; 184:1877-1879.
17. Singer K, Chernoff AI, Singer L. Studies on abnormal hemoglobins: Further variations in sickle cell anemia and other hematologic disorders by means of alkali denaturation. *Blood* 1951; 6: 413-416.
18. Schleider CTH, Mayson SM, Huisman THJ. Further modification of the microchroma tographic determination of hemoglobin A₂. *Hemoglobin* 1977; 1: 503-507.
19. Poncz M, Sdowiejezyk D, Harpel B, *et al.* Construction of human gene libraries from small amounts of peripheral blood: Analysis of beta like globin genes. *Hemoglobin* 1982; 6: 27-40.
20. Newton CR, Graham A, Hiptinstall LE, *et al.* Analysis of any point mutation in DNA. The amplification refractory mutation system (ARMS). *Nuc Acid Res* 1989; 17: 2503-2516.
21. Varawalla NY, Old JY, Sarkar R, *et al.* The spectrum of (3-thalassemia mutations on the Indian subcontinent: The basis for prenatal diagnosis. *Br J Haem* 1991; 78: 242-245.
22. Agarwal S, Naveed M, Gupta UR, *et al.* Characterization of (3-thalassemia mutations in 57 fi-thalassemia families seen at Lucknow. *Indian J Med Res* 1994; 100: 106-111.
23. Fucharoen S, Winichagoon P. Hemoglobinopathies in South East Asia. *Hemoglobin* 1987; 11: 65-70.
24. Bunn HF, Forget BG. *In: Hemoglobin: Molecular, Genetic and Clinical Aspects.* Philadelphia, W.B. Saunders Company, 1986; pp 322-379.
25. Embury SH, Kropp GL, Stanton TS, *et al.* Detection of the HbE-mutation using the color complementation assay: Amplification to complex genotyping. *Blood* 1990; 76: 619-623.
26. Goossens M, Dozy AM, EMbuzy SM, *et al.* Triplicated alpha-gene loci in humans. *Proc Natl Acad Sci USA* 1980; 77: 518-521.
27. Garewal G, Fearon CW, Warren TC *et al.* The molecular basis of p-thalassemia in Punjabi and Maharashtrian Indians includes a multilocus etiology involving triplicated alpha-globin loci. *Br J Hem* 1994; 86: 372-376.
28. Gupta RB, Tiwary RS, Pande PL, *et al.* Hemoglobinopathies among the Gond Tribal groups of Central India. Interaction of alpha and P-thal with (3-chain variants. *Hemoglobin* 1991; 15: 441-450.

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