
Original Articles

HAEMOPHILUS INFLUENZAE TYPE B VACCINE IN INDIA: NEED AND TIMING, IMMUNOGENE CITY AND TOLERANCE

**Debyani Acharya, Sheila Bhawe, Vaishali Joshi,
Ashish Bavdekar and Anand Pandit**

From the Department of Pediatrics, K.E.M. Hospital, Pune 411 011.

Reprint requests: Dr. Sheila Bhawe, Associate Consultant in Pediatric Research, Department of Pediatrics, KEM Hospital, Pune 411011.

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Objective: (i) To assess the natural immunity and susceptibility to *Haemophilus influenzae* type b (Hib) infections in children in India, (ii) To study the immunogenicity and tolerance of Hib vaccine (ACTHIB) in young infants. **Designs:** (i) Cross sectional study, (ii) Prospective trial. **Setting:** Well baby and immunization clinics. **Methods:** (i) PRP antibody titers against Hib estimated in 172 healthy infants and children aged 1 month to 10 years, (ii) Antibody titres estimated before and after ACTHIB vaccine given with primary immunization (age group 6 to 8 weeks) in 50 babies. **Results:** (i) Naturally protective levels of Hib antibodies found in less than 20% of infants under one year, but in over 80% above 4 years, (ii) Seroconversion after ACTHIB vaccination was 100% with very high protective levels. There were no significant adverse reactions. **Conclusions:** ACTHIB vaccine proved to be safe and highly immunogenic. As susceptibility to Hib is highest in the first year of life, the vaccine should be recommended in the primary immunization schedule (combined with DPT). The very high titers achieved suggest the possibility of decreasing the number of doses or the amount of antigen to reduce the prevalent high cost.

Key words: *Haemophilus influenzae* infection, Immunization.

Haemophilus influenzae type 'b' (Hib) is a common cause of invasive bacterial infections in children aged 3 months to 5 years, causing a spectrum of serious illnesses such as meningitis, epiglottitis and pneumonia(1,2). Mortality and morbidity of these conditions is high, especially if treatment is delayed(3). Emerging resistant strains pose further problems in successful treatment⁴). High titers of anti Poly Ribosyl Phosphate (PRP) antibodies in convalescent sera led to the development of conjugate vaccines, which since 1988, are in

regular use in developed countries(5-7). Routine vaccination in these countries has led to a remarkable decline in the incidence of Hib infections(8,9). In Finland, for example, the incidence in children under 5 years has fallen from 52/100,000 in pre-vaccination era to virtually nil since 1992(7).

The exact incidence of Hib, related disease in children in India is largely unknown. The few reported studies quoting 8 to 14% of meningitis(10-12) and 7 to 15% of

lobar pneumonias are likely to be underestimated because of poor bacterial culture facilities in our laboratories(13). Though effective, Hib conjugate vaccines are expensive and not yet available for routine use in India. A preliminary multicentric trial of the vaccine in 125 Indian children between the ages of 18 to 24 months (1st booster age group) has given encouraging results(14). But before recommending routine immunization against Hib in our country the questions that need to be answered are: (i) What is the natural prevalence of Hib in children in India? Is the vaccine really needed in our country?; (ii) What is the critical period of susceptibility to the disease and therefore what is the optimum timing of the vaccine?; and (iii) What is the immunogenicity and tolerance of the ACTHIB vaccine in combination with DPT in young infants? This study was specifically planned to address the aforementioned issues.

Subjects and Methods

These studies were conducted by the Department of Pediatrics, KEM Hospital, Pune, over a period of one year. The protocol was reviewed and approved by the Ethics Committee of the hospital.

Subjects

Study I: Study of Cross Sectional Survey of Anti-PRP Antibodies .

One hundred and seventy two healthy children between the ages of one month to 10 years attending the Well Baby Clinic or Immunization Clinic were randomly selected in their respective age groups. Children with acute infections and those suffering from chronic debilitating illnesses, were excluded.

As anticipated population prevalence in the country is unknown, it was assumed to be 50%. The estimated prevalence on 170 children will fall within 7.5 percentage

points of the true prevalence with 95% confidence.

Study II: Immunogenicity and Tolerance Study

Fifty infants of age 6 to 8 weeks and requiring primary schedule of DPT and polio vaccination under Universal Immunization Programme (UIP) were recruited. Babies suffering from any infection, neurologic disorders, immunocompromized babies, or those undergoing steroid therapy were excluded. Informed consent was obtained from parents of the babies after giving full description of the vaccine and schedule of blood collection.

Vaccine

ACTHIB (Pasteur Merieux) is a capsular polysaccharide covalently conjugated to tetanus protein (PRP-T). The 0.5ml dose of reconstituted vaccine corresponds to 10 meg of polysaccharide. DPT vaccine and Polio Vaccine (OPV) were supplied through UIP programme. Vaccines were maintained in cold chain conditions.

Vaccination Schedule

Study II (a): The babies recruited were randomly allocated to Groups A or B. Babies enrolled in Group A were given 0.5 ml ACTHIB intramuscularly (lateral region of thigh), in association with DPT, *i.e.*, they received the DPT at a different site. Babies enrolled in Group B received combined vaccination with DPT, *i.e.*, ACTHIB and DPT were mixed extemporaneously in the same syringe and administered intramuscularly. The vaccination was carried out at approximately 2,3, and 4 months of age. At the same time, they also received the OPV as per the primary vaccination schedule.

Blood Collection

In Study I, 3 ml of blood was collected from all the enrolled children by venepuncture at one time. In Study II, 3 ml of

blood was collected prior to vaccination and four weeks after the last dose of vaccination from all enrolled babies. Sera were separated by centrifugation and coded.

Adverse Reactions

The babies who received vaccination were observed for any immediate adverse reactions upto 1 hour after vaccination. Parents were instructed to record and report local as well as systemic reactions such as fever, irritability, persistent crying, anorexia, vomiting, rash or convulsions.

Serological Analysis

All the separated sera were carefully stored at -20°C and were dispatched in frozen state to Lyon, France. Serum anti-PRP antibody was measured with a FARR type of FJA using intrinsically labelled PRP that used ^{125}I labelled polysaccharide(15).

Titers above 0.15 mcg/ml were considered as seroconversion (natural protection threshold) and whereas, levels above 1

mcg/ml taken to indicate long term vaccine protection threshold(16,17).

Statistics

Postvaccination geometric mean antibody titers (GMT) between the two study groups were analyzed by unpaired 't' test. Pre and post vaccination titers within each group were analyzed using paired 't' test.

From our data and sample size, the power of the study for estimating the immunogenicity of the vaccine given combined or associated with DPT vaccine exceeds 0.9. Power calculations have been made considering two tailed distribution and 95% level of significance.

Results

Study I: In the cross-sectional survey a total of 172 samples were collected. Two samples could not be analyzed due to insufficient quantity. The analysis of 170 samples shows age related increase in anti-PRP antibodies (*Table I*). Irrespective of age group,

TABLE I—Cross-sectional Survey of Anti PRP Antibodies (Against Hib) in 170 Healthy Infants and Children.

Age group	n (n males)	n (%) with Anti PRP level >0.15 mcg/ml	Mean anti PRP (\pm SE)
Group I (1-12 mo)	27 (21)	5 (19)	0.30(0.08)
Group II (12-24 mo)	25 (14)	9 (36)	0.21(0.03)
Group III (24-36 mo)	23 (18)	8 (35)	0.30(0.07)
Group IV (36-48 mo)	9 (6)	6 (67)	0.55(0.35)
Group V (48-60 mo)	16 (10)	13 (81)	0.94(0.50)
Group VI (>60 mo)	70 (36)	60 (86)	1.18(0.31)
Total	170	101 (59.4)	

levels above 0.15 mcg/ml (natural protection) were observed in 101 babies of which 16 had titers above 1 mcg/ml (long term protection). Two one month old babies had titers of 0.57 and 1.3 mcg/ml, possibly due to transplacental transfer. However, 22 (81%) babies below the age of 12 months had titers below 0.15 mcg/ml, *i.e.*, they were susceptible to Hib infections. The susceptibility reduced with increasing age so that less than 30% of children over the age of 3 years had titers below 0.15 mcg/ml.

Study II: All 50 babies completed the primary schedule of vaccination and the subsequent follow up. Paired sera were available from 48 babies as two refused postvaccination blood sampling. There was no significant difference in the mean weight and age at initiation of study in the two groups.

Table II shows immunogenic response of ACTHIB in primary immunization schedule. The pre vaccination anti PRP antibodies were higher than 0.15 mcg/ml (natural protection) in only three babies in Group A and seven babies in Group B. The post vaccination titers were significantly higher in both the groups with 100% seroconversion in all ($p = 0.0001$). There was no significant difference in the sero-

conversion rate of babies with prevac titers below or above 0.15 µg/ml. All but one baby achieved post vaccination titers of more than 1 mcg/ml (long term vaccine induced protection). The post vaccination titers in Group B (*i.e.*, DPT and ACTHIB combined in same syringe) were significantly higher than in Group A (ACTHIB and DPT at different sites) ($p = 0.003$).

Adverse Reactions

There were no serious adverse effects in the form of vomiting, convulsions or hypotonia. A total of 11 babies had mild fever lasting for 48 hours. Local pain and erythema were seen in 2 babies and one baby developed induration, which subsided on its own without any surgical intervention. There were no differences in the groups receiving ACTHIB and DPT concurrently or in combination.

Discussion

Diseases known to be related to Hib infections such as meningitis and pneumonias are not uncommon in India(10-13). Mortality and morbidity of these conditions is high especially in infants and children under 3 years(18,19). Our cross-sectional survey for Hib antibodies con-

TABLE II—Anti PRP Antibody Titers (µg/ml) in 48 children given with Primary DPT.

Anti PRP antibodies (µg/ml)	Group A (24)		Group B (24)	
	Associated	Injections	Combined	Injections
	Prevac	Postvac	Prevac	Postvac
n (%) < 0.15	21 (88)	0 (0)	17 (71)	0 (0)
n (%) > 0.15	3 (12)	24 (100)	7 (29)	24 (100)
n (%) > 1.0	0 (0)	23 (96)	0 (0)	24 (100)
GMT	0.17	11.97	0.21	31.48
(SEM)	(1.09)	(1.3)	(1.13)	(1.2)

Prevac and postvac GMT in Group A – $p < 0.0005$
 Prevac and postvac GMT in Group B – $p < 0.0005$
 Postvac GMT in Group A and Group B – $p < 0.003$

ducted in 170 healthy children in various age groups (1 month to 10 years) demonstrates a high prevalence of subclinical Hib infections, and least natural protection under the age of 3 years, especially under one year.

More than 70% of children above the age of 3 years had natural protective antibody titers above 0.15 mcg/ml indicating subclinical infections. However, 70% of infants and children under age 3 years had titers below 0.15 mcg/ml and were hence, susceptible to Hib infections. This susceptibility was highest (81%) under the age of 1 year. This study therefore, emphasizes the need for Hib vaccination in our country and that too at an earlier age, namely, in the primary immunization age group rather than at booster age of 18 months as given in our recently reported multicentric study(14).

The vaccine ACTHIB (Pasteur Merieux) proved highly successful with 100% seroconversion with antibody titers well above 1 mcg/ml in all but one baby. Titers of more than 1 mcg/ml are generally correlated with long term protection(20). Infact, the post vaccination titers with 3 doses in our study, were two to three times greater than generally reported in western countries(21-23). Similar strikingly high post immunization response is also reported in Venezuelan children(24). The high antibody response in developing countries could well be due to subclinical infections or racial variations.

The enhanced serological conversion in our babies suggest the possibility of administering fewer doses of vaccine or smaller amounts of antigen with great potential for saving public health resources. This hypothesis needs to be corroborated with further studies of immune response following each dose to formalise the optimum schedule in our country.

Immunogenicity and safety of PRP-T (ACTHIB) given in combination with DPT has been assessed in several studies besides ours. The antibody response of other antigen has not been affected with ACTHIB(25,26). In our study, the antibody response in babies receiving PRP-T vaccination in combination with DPT was infact better than when given alone or at different sites.

The vaccine was remarkably well tolerated as seen in our earlier study (booster age group)(14). There were no significant differences in the local or systemic reactions in the two study groups. The safety and efficacy of Hib vaccine has been confirmed by other studies earlier(22).

In conclusion, the high susceptibility of our infants and young children to Hib infections emphasizes the urgent need for a protective vaccine in India. The vaccine must be introduced early in life, preferably in a conjugate preparation combining DPT with Hib along with OPV. However, prospective studies of suspected infections are necessary to determine the exact incidence of Hib in India.

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REFERENCES

1. Broome C V. Epidemiology of *Haemophilus influenzae V* in US. *Pediatr Infect Dis* 1987, 6: 779-782.
2. Bijlmer HA. Worldwide epidemiology of *Haemophilus influenzae* meningitis, Industrialized versus non industrialized countries. *Vaccine* 1991, 9 (Suppl): S5 -S9.

3. Cochi S.L., Broome CV, Highwater AW. Immunization of US children with *H. influenzae* type b polysaccharide vaccine: A cost effectiveness model of strategy assessment. JAMA 1985, 253: 521-529.
4. Peltola H, Rod TO, Jonsdottir K, Bottiger M, Coolidge AS. Life threatening *H. influenzae* in Scandinavia. Analysis of incidence of bacteriological characteristics. Rev Inf Disease 1990,12: 708-715.
5. Kathy H, Peltola H, Karnako V, *et al.* The protective level of serum antibodies to the capsular polysaccharide of *Haemophilus influenzae* type b. J Infect Dis 1983, 147: 1100-1103.
6. AAP Committee on Infectious Disease. *H. influenzae* vaccine. Update Pediatrics 1989, 84: 386-387.
7. Peltola H, Kilpi T, Anuila M. Rapid disappearance of *Haemophilus influenzae* meningitis after immunization. Lancet 1992, 340: 592-594.
8. Adams WG, Deaver KA, Cochi SI, *et al.* Decline in childhood *Haemophilus influenzae* type b (Hib) disease in the Hib vaccine era. JAMA 1983, 269: 221-226.
9. Murphy TV, White KE, Pastor P, *et al.* Declining incidence of *Haemophilus influenzae* type b disease since introduction of vaccination. JAMA 1993, 269: 246-248.
10. Achar ST, Thambiah S. Pyogenic meningitis in Indian children. Indian J Child Health 1954, 3: 5-8.
11. Vishnu Bhatt B, Verma IC, Puri RK, *et al.* Profile of pyogenic meningitis. J Indian Med Assoc 1991, 89: 224-226.
12. Tamaskar V, Bhandari NR. A clinico bacteriological study of meningitis in childhood. Indian J Pediatr 1976, 43: 226-230.
13. Kumar L, Ayyagari A. The etiology of lobar pneumonia and emphysema thoracics in children. Indian Pediatr 1984, 21: 133-138.
14. Nanavaty N, Pandit AN, Acharya DK, *et al.* Evaluation of immunogenicity tolerance of single dose of PRP-T vaccine. Indian Pediatr 1995, 32:1077-1081.
15. Kuo JSC, Monji N, Schwalbe RS, *et al.* A radioactive antigen binding assay for measurement of antibody to Hib polysaccharide. J Immunol Methods 1981, 43: 35-47.
16. Robbins JB, Parke JC, Schneerson R, Whisnant JK. Quantitative measurement of "natural" and immunization induced *Haemophilus influenzae* type b capsular polysaccharide antibodies. Pediatr Res 1973, 7:103-110.
17. Kayhty H, Peltola H, Karanko V, Makela PH. The protective level of serum antibodies to the capsular polysaccharide of *Haemophilus influenzae* type b. J Infect Dis 1983,147:1100.
18. Todd JK, Bruhn FW. Severe *Haemophilus influenzae* infections: spectrum of disease. Am J Dis Child 1975,129: 607-611.
19. Klein JO, Feigin RD, McCracken GH. Report of the task force on diagnosis and management of meningitis. Pediatrics 1986, 78 (Suppl): 959-982.
20. Claesson BA, Schneerson R, Robbins JB, *et al.* Protective levels of serum antibodies stimulated in infants by two injections of *Haemophilus influenzae* type b capsular polysaccharide tetanus toxoid conjugate. J Pediatr 1989,114: 97-100.
21. Decker MD, Edwards KM, Bradley R, Palmer P. Comparative trial in infants of four conjugate *Haemophilus influenzae* type b vaccine. J Pediatr 1992,120: 184-189.
22. Fritzell B, Plotkin S. Efficacy and safety of a *Haemophilus influenzae* type b capsular polysaccharide tetanus protein conjugate vaccine. J Pediatr 1992,121: 355-362.
23. Greenberg DP, Vadheim CM, Partridge S *et al.* Immunogenicity of *Haemophilus influenzae* type b tetanus toxoid conjugate vaccine in infants. J Inf Dis 1994, 170: 76-81.
24. Casillo De Febres O, Decker MD, Estopinan M, Bordonas G, Edwards K.

- Enhanced antibody response in Venezuelan infants immunized with *Haemophilus influenzae* type b tetanus toxoid conjugate vaccine. *Pediatr Infect Dis J* 1994, 13: 635-639.
25. Watterberg N, Dagan R, Arbelli Y, *et al.* Safety and immunogenicity of *Haemophilus influenzae* type b-tetanus protein conjugate vaccine mixed in the same syringe with diphtheria-tetanus pertussis vaccine in young infants. *Pediatr Infect Dis J* 1991, 10: 758-763.
26. Eskola K H, Gordon LK, Hovi T, *et al.* Simultaneous administration of *Haemophilus influenzae* type b capsular polysaccharide diphtheria toxoid conjugate vaccine with routine diphtheria-pertussis-tetanus and inactivated poliovirus vaccinations of childhood. *Pediatr Infect Dis J* 1988, 7: 480-484.
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